



Effects of propofol, desflurane, and sevoflurane on respiratory functions following endoscopic endonasal transsphenoidal pituitary surgery: a prospective randomized study

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Background: General anesthesia with intravenous or inhalation anesthetics reduces respiratory functions. We investigated the effects of propofol, desflurane, and sevoflurane on postoperative respiratory function tests.

Methods: This single-center randomized controlled study was performed in a university hospital from October 2015 to February 2017. Ninety patients scheduled for endoscopic endonasal transsphenoidal pituitary surgery were randomly categorized into either of these three groups: propofol (n = 30, the Group TIVA), desflurane (n = 30, the Group D) or sevoflurane (n = 30, the Group S). We analyzed the patients before, after, and 24 h following surgery, to identify the following parameters: forced expiratory volume in 1 second (FEV₁) %, forced vital capacity (FVC) %, FEV₁/FVC, and arterial blood gases (ABG). Furthermore, we also recorded the intraoperative dynamic lung compliance and airway resistance values.

Results: We did not find any significant differences in FEV₁ values (primary outcome) among the groups (P = 0.336). There was a remarkable reduction in the FEV₁ and FVC values in all groups postoperatively relative to the baseline (P < 0.001). The FVC, FEV₁/FVC, ABG analysis, compliance, and airway resistance were similar among the groups. Intraoperative dynamic compliance values were lower at the 1st and 2nd hours than those immediately after intubation (P < 0.001).

Conclusions: We demonstrated that propofol, desflurane, and sevoflurane reduced FEV₁ and FVC values postoperatively, without any significant differences among the drugs.

Keywords: Airway resistance; Desflurane; Lung compliance; Propofol; Respiratory function tests; Sevoflurane.

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Introduction

General anesthesia associated changes in respiratory functions may be attributed to reduced lung compliance, closed airway, and lower functional residual capacity that may be etiologically associated with the formation of atelectasis. Halogenated anesthetics-induced loss of respiratory muscle tone can lead to formation of atelectasis [1]. Similarly, a single bolus dose of propofol may reduce twitch diaphragmatic pressure. Animal studies have demonstrated that inhalation anesthetics impaired hypoxic pulmonary vasoconstriction (HPV) and increased shunt fraction, whereas propofol does not affect HPV [2–5]. Considering this, propofol and inhalation anesthetics are expected to demonstrate different effects on pulmonary functions. However, each inhalation anesthetic affects the respiratory system differently; for instance halothane, sevoflurane, and isoflurane relax the airways by depleting sarcoplasmic reticulum Ca^{2+} stores, unlike desflurane [6–8].

Several studies have previously compared the effect of propofol and inhalation anesthetics on lung functions in different surgical procedures, except for intracranial surgeries. Tiefenthaler et al. [9] and Zoremba et al. [10] reported that propofol impaired lung functions more than inhalation anesthetics. Alternatively, no significant difference was found in other studies [11,12]. Proper titration of the positive end expiratory pressure level according to lung compliance may reduce the side effects of anesthetic drugs on lung function. However, positive end expiratory pressure (PEEP) titration in neurosurgical procedures could be problematic due to the complications associated with increased intracranial pressure. Therefore, to compare the effects of propofol, desflurane, and sevoflurane on respiratory functions we selected endoscopic endonasal transsphenoidal pituitary surgery patients who were already on a suboptimal ventilation with fixed PEEP levels.

Therefore, we aimed to compare the effects of three agents on forced expiratory volume in 1 second (FEV_1), forced vital capacity (FVC), lung compliance, and airway resistance during the postoperative 24 hours in patients who were undergoing endoscopic endonasal transsphenoidal pituitary surgery.

Materials and Methods

Study design

The Ethics Committee of Cerrahpasa School of Medicine, Istanbul, Turkey (Chairperson Prof Ozgur Kasapcopur) provided ethical approval for this study on 02 June 2015 (Ethical Committee No NAC 83045809/604.01/02). This study was registered to “clinicaltrials.gov with the number NCT02709863”. This prospective, randomized, and parallel study was performed

between October 2015 and February 2017 in Istanbul University- Cerrahpasa, Cerrahpasa School of Medicine, Neurosurgical Operation Rooms and Neurosurgery ward.

Participants

Ninety American Society of Anesthesiologists physical status classification (ASA) I–II patients, aged between 18 to 70 years, who provided their written informed consent and were scheduled for elective endoscopic endonasal transsphenoidal pituitary surgery were included in the study. Patients presenting with any of the following conditions were excluded from the study: neurological disorders hindering the communication, obstructive or restrictive lung disease, heart failure, liver failure, kidney failure, smoking, drug or alcohol addiction, and dementia. Furthermore, patients who developed bronchospasm or laryngospasm or remained unconscious at the end of the surgery were excluded from the study.

Each patient underwent numerous respiratory functions tests (Spirolab III COLOUR LCD) and arterial blood gases (ABG) analyses (Cobas b 221) with the patient placed in the Fowler position before administration of anesthesia (Baseline). Three acceptable spirometry tests were acquired and the best measurement was recorded. We subsequently recorded hemodynamic parameters, forced expiratory volume in 1 second (FEV_1) %, forced vital capacity (FVC) % and FEV_1/FVC , pH, arterial partial pressure of O_2 (PaO_2), arterial partial pressure of CO_2 ($PaCO_2$), bicarbonate concentration ($[HCO_3^-]$), base excess (BE), and PaO_2/FiO_2 .

Randomization

Patients were categorized using a computer-generated randomization method, by using numbered sealed envelopes to one of the following three groups by the anesthesiology nurse: total intravenous anesthesia (the Group TIVA, $n = 30$), desflurane (the Group D, $n = 30$), or sevoflurane (the Group S, $n = 30$) based general anesthesia.

Blinding

The on-duty anesthesiologists enrolled the participants, while another technician, who was blinded to the anesthesia method, performed the postoperative respiratory function tests.

Interventions

Patients were sedated with intravenous midazolam (0.03 mg/kg) before the surgery in the anesthesia preparation room. In the operating room, after routine monitoring, bispectral index

(BIS) and train-of-four (TOF) were monitored. Anesthesia was induced with propofol (1.5–2 mg/kg), rocuronium (0.5 mg/kg), and remifentanyl (0.1 µg/kg/min). After 3 minutes of manual ventilation with oxygen/air ($FiO_2 = 0.8$) patients were intubated with 7.5 mm internal diameter endotracheal tube for women and 8.0 mm internal diameter endotracheal tube for men. The cuff was inflated with air, and cuff pressure was maintained at 25 cmH₂O. Patients were ventilated using a volume-controlled mode, tidal volume: 8 ml/kg (ideal body weight), $FiO_2 = 0.4$, inspiration:expiration ratio of 1:2, PEEP: 5 cmH₂O and the respiratory rate (9–12 /min) was adjusted to maintain PaCO₂ in the range of 36 to 38 mmHg. Anesthesia was maintained with remifentanyl (0.05–0.15 µg/kg/min) and rocuronium (0.3 mg/kg/h), followed by infusion of propofol (4 to 8 mg/kg/h), desflurane (1 minimum alveolar concentration [MAC] in oxygen/air), or sevoflurane (1 MAC in oxygen/air). Propofol and inhalation anesthetic concentrations were adjusted to maintain the BIS range between 45 and 60.

Intraoperative dynamic lung compliance (C) and airway resistance (R_{aw}) were recorded after intubation and at 1st and 2nd hour. ABG analysis was repeated after orotracheal intubation, 15 min before extubation, and 30 min and 24 h after surgery. Hemodynamic parameters and respiratory rates were recorded at the same intervals.

The right radial artery and urinary catheters were placed. Surgical incision site was infiltrated with a maximum of 20 ml of lidocaine 1% and adrenaline (1 : 100.000) mixture. All patients

were operated in the supine position. Intraoperative analgesia was maintained with an infusion of remifentanyl and morphine. The maximum infusion dose of remifentanyl was 0.20 µg/kg/min to intraoperatively control hemodynamic response. If it could not be controlled with remifentanyl alone, we administered intravenous morphine with the maximum dose 0.1 mg/kg. Patients received ondansetron (4 mg) as an antiemetic prophylaxis at the end of the surgery. Residual muscle relaxation was reversed with sugammadex (2 mg/kg) at the end of surgery. The endotracheal tube was removed when the train of four ratio was 0.9 and the patient regained consciousness. The patient was subsequently transferred to the recovery room. All patients had previously been instructed about the visual analogue scale (VAS) from 0 to 10, with 0 representing no pain and 10 indicating the worst pain imaginable. Intravenous morphine (2 mg) administration was provided and repeated every 10 minutes if the VAS score was higher than 3. Total morphine consumption and duration of surgery were recorded. Patients were transferred to the ward when the modified Aldrete score was more than 9.

The respiratory function tests for each were repeated 30 min and 24 h postoperatively.

Outcomes

The primary endpoint of the present study was to compare the effects of propofol, desflurane, and sevoflurane on the FEV₁, while the secondary endpoints were to compare their effects on

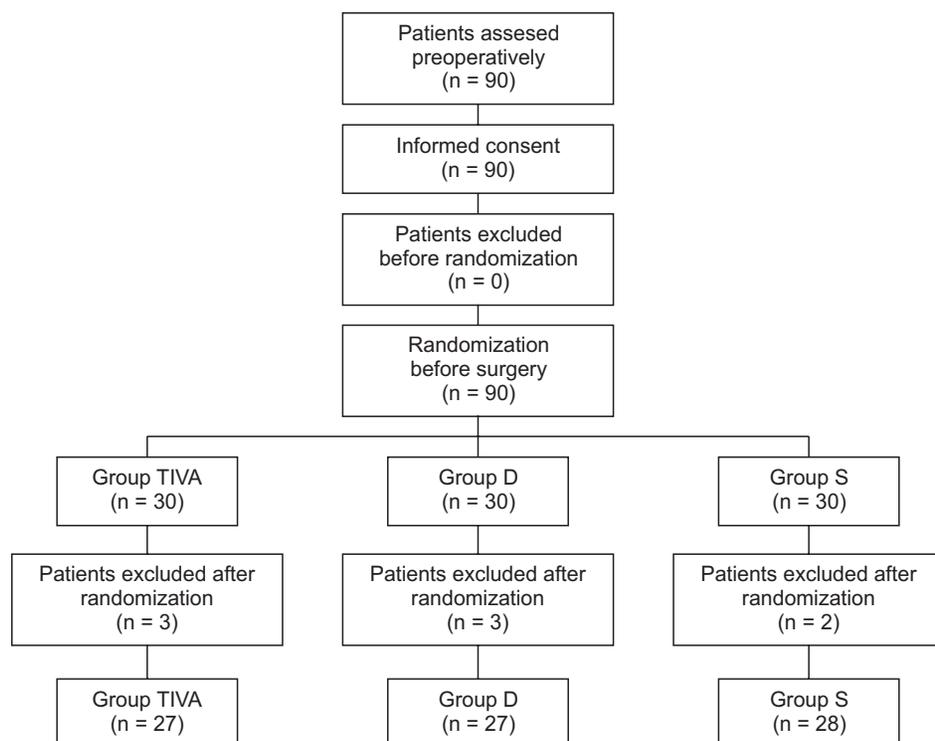


Fig. 1. Flow of participants in the study.

the FVC, FEV₁/FVC, ABG analysis, dynamic compliance, and airway resistance in patients undergoing endoscopic endonasal transsphenoidal pituitary surgery during the postoperative 24 h.

Statistical analysis and sample size

According to Tiefertal et al. [9], 27 patients were needed in each group to detect a minimum inter-group difference of 20% with regard to FEV₁ values, with a probability of error type II of 20% ($\beta = 0.2$) and error type I of 1% ($\alpha = 0.01$).

All data were expressed as a number or mean \pm SD. The statistical package for the social sciences (SPSS) 15.0 (SPSS Inc, Chicago) was used for statistical analysis. Pearson χ^2 test was used to compare inter-group qualitative variables, such as gender and ASA, which demonstrated binary change. The Shapiro Wilk normality test was used to evaluate the distribution of data.

The One-Way ANOVA was used to compare normally distributed variables among the groups. The Kruskal Wallis analysis was used to compare the variables that were not normally distributed. The Mann Whitney *U* test with Bonferroni correction was performed to highlight the inter-group differences when the differences were found. Statistical significance was determined via adjusted alpha ($= 0.05/3 = 0.01666$) by Bonferroni correction. The repeated measures ANOVA was used for intra- and inter-group comparisons; furthermore, $P < 0.05$ and $P < 0.016$ were considered as indicators of statistical significance, respectively.

Results

We had initially included 90 patients in this study. Three patients from Group TIVA were excluded from the study due to

Table 1. Patient Characteristics

	Group TIVA (n = 27)	Group D (n = 27)	Group S (n = 28)	P value
ASA (I/II) [†]	8/19	13/14	6/22	0.115
Gender (M/F) [†]	15/12	12/15	12/16	0.641
Age (yr) [†]	43.1 \pm 13.0	46.5 \pm 13.1	45.2 \pm 12.5	0.615
Height (cm) [§]	168.7 \pm 9.2	166.6 \pm 9.1	168.0 \pm 6.9	0.494
Weight (kg) [†]	85.4 \pm 16.4	80.5 \pm 16.7	84.6 \pm 16.4	0.510
BMI (kg/m ²) [†]	29.8 \pm 5.0	28.5 \pm 6.0	29.8 \pm 5.7	0.633
Duration of surgery (min) ^{§,}	180.0 \pm 16.9	195.1 \pm 8.4	173.1 \pm 5.9	< 0.001*

Values are presented as numbers or mean \pm SD. ASA: American Society of Anesthesiologists, BMI: body mass index, n: number. Comparison among the groups, *Indicates a statistically significant difference ($P < 0.016$). [†]Pearson Chi-Square, [‡]Oneway ANOVA, [§]Kruskal Wallis, ^{||}Bonferoni corrected Mann Whitney *U*.

Table 2. Results of Pulmonary Function Test

	Group TIVA (n = 27)	Group D (n = 27)	Group S (n = 28)	P value
FEV ₁ (%)				0.336*
Baseline	81.6 \pm 19.0	83.4 \pm 18.9	73.4 \pm 17.8	
30th min	63.8 \pm 22.0	67.5 \pm 20.3	60.40 \pm 22.3	
24 h	71.4 \pm 22.8	69.03 \pm 17.5	67.96 \pm 16.9	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
FVC (%)				0.031*
Baseline	84.0 \pm 21.4	88.6 \pm 27.4	71.9 \pm 18.7	
30th min	65.6 \pm 20.4	73.5 \pm 26.3	62.2 \pm 22.2	
24 h	74.0 \pm 22.9	69.4 \pm 14.8	66.7 \pm 17.1	
P value [†]	< 0.001 [†]	< 0.001 [†]	0.002 [†]	
FEV ₁ /FVC				0.104*
Baseline	102.0 \pm 16.3	99.0 \pm 19.0	107.9 \pm 16.2	
30th min	101.7 \pm 15.6	101.5 \pm 17.7	100.9 \pm 14.1	
24th h	100.0 \pm 15.1	105.4 \pm 17.7	105.6 \pm 11.7	
P value [†]	0.815	0.243	0.009 [†]	

Values are presented as mean \pm SD. FEV₁: Forced expiratory volume in the 1st second, FVC: Forced vital capacity. *comparison among the groups, Repeated Measures of ANOVA. No difference in FEV₁ ($P = 0.336$), FVC ($P = 0.031$), and FEV₁/FVC among the groups ($P = 0.104$) ($P < 0.016$ indicates statistically significant difference among the groups). [†]within group comparisons, Repeated Measures of ANOVA. Reduction in the FEV₁ and FVC values in all groups at the postoperative period than the baseline ($P < 0.001$ and 0.002 , respectively) ($P < 0.05$ indicates statistically significant difference within group comparisons).

the following reasons: one developed bronchospasm necessitating switching from propofol infusion to sevoflurane inhalation, and two refused to postoperatively repeat respiratory function

tests. Three patients from Group D were excluded, since one developed hematoma at the surgical site intraoperatively and had to be admitted to the intensive care unit, one did not adequately

Table 3. Results of Arterial Blood Gas Analysis

	Group TIVA (n = 27)	Group D (n = 27)	Group S (n = 28)	P value
PaO ₂ (mmHg)				0.299*
Baseline	100.5 ± 19.4	94.7 ± 16.4	91.7 ± 13.7	
After intubation	140.4 ± 41.3	151.5 ± 53.8	143.5 ± 32.8	
Before extubation	153.9 ± 38.6	159.2 ± 40.6	160.8 ± 29.3	
30th min	121.0 ± 35.0	126.2 ± 38.8	132.1 ± 46.2	
24 h	93.7 ± 20.9	83.7 ± 16.8	84.6 ± 16.1	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
PaCO ₂ (mmHg)				0.843*
Baseline	34.1 ± 5.0	34.0 ± 3.6	35.8 ± 3.1	
After intubation	35.3 ± 3.2	34.9 ± 3.6	35.4 ± 4.7	
Before extubation	37.1 ± 2.6	36.4 ± 2.7	37.1 ± 3.2	
30th min	42.0 ± 4.0	40.9 ± 3.4	42.5 ± 3.4	
24 h	35.3 ± 4.5	35.4 ± 3.8	35.4 ± 3.5	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
pH				0.632*
Baseline	7.4 ± 0.04	7.4 ± 0.02	7.4 ± 0.03	
After intubation	7.4 ± 0.03	7.4 ± 0.04	7.4 ± 0.05	
Before extubation	7.4 ± 0.03	7.4 ± 0.03	7.3 ± 0.04	
30th min	7.3 ± 0.03	7.3 ± 0.03	7.3 ± 0.04	
24 h	7.4 ± 0.04	7.4 ± 0.04	7.4 ± 0.03	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
SaO ₂ (%)				0.736*
Baseline	97.7 ± 1.2	97.5 ± 1.2	97.5 ± 1.0	
After intubation	98.6 ± 0.7	98.2 ± 1.5	98.8 ± 0.5	
Before extubation	98.8 ± 0.3	98.4 ± 0.8	98.9 ± 0.6	
30th min	97.8 ± 1.6	98.0 ± 1.4	98.1 ± 1.6	
24 h	97.2 ± 1.8	96.9 ± 1.6	97.0 ± 1.6	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
HCO ₃ ⁻ (mmol/L)				0.685*
Baseline	24.0 ± 2.0	23.5 ± 1.3	23.2 ± 1.3	
After intubation	24.4 ± 1.4	23.5 ± 1.4	23.2 ± 2.2	
Before extubation	24.2 ± 1.3	23.4 ± 1.6	23.3 ± 1.8	
30th min	24.1 ± 1.5	23.1 ± 1.3	23.3 ± 1.9	
24 h	25.3 ± 1.6	24.6 ± 1.7	24.9 ± 1.9	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
BE (mmol/L)				0.698*
Baseline	-0.7 ± 2.2	-1.4 ± 1.5	-1.4 ± 1.4	
After intubation	-0.3 ± 1.6	-1.3 ± 1.6	-1.5 ± 2.3	
Before extubation	-0.3 ± 1.4	-1.3 ± 1.7	-1.4 ± 2.0	
30th min	-0.4 ± 1.8	-1.2 ± 1.5	-1.1 ± 2.2	
24 h	0.7 ± 1.9	0.07 ± 1.9	0.5 ± 2.2	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
Lactate (mmol/L)				0.027*
Baseline	1.5 ± 0.6	1.1 ± 0.4	1.4 ± 0.6	
After intubation	1.5 ± 0.5	1.2 ± 0.5	1.5 ± 0.6	
Before extubation	1.4 ± 0.6	1.2 ± 0.5	1.7 ± 0.9	
30th min	1.4 ± 0.5	1.4 ± 0.6	1.8 ± 0.9	
24 h	1.5 ± 0.6	1.4 ± 0.8	1.3 ± 0.6	
P value [†]	0.723	0.051	0.012 [†]	

Table 3. Continued

	Group TIVA (n = 27)	Group D (n = 27)	Group S (n = 28)	P value
PaO ₂ /FiO ₂				0.268*
Baseline	457.4 ± 78.7	443.0 ± 90.7	437.1 ± 65.1	
After intubation	349.4 ± 101.1	376.6 ± 138.1	358.2 ± 82.4	
Before extubation	380.9 ± 100.2	390.4 ± 111.10	401.4 ± 73.6	
30th min	322.5 ± 81.9	326.9 ± 99.8	339.7 ± 113.9	
24th h	425.0 ± 87.8	378.4 ± 83.9	392.7 ± 67.8	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	

Values are presented as mean ± SD. PaO₂: Partial pressure of arterial oxygen, PaCO₂: Partial pressure of arterial carbon dioxide, pH: Potential of hydrogen, SaO₂: Arterial oxygen saturation, [HCO₃⁻]: Bicarbonate, BE: Base excess, PaO₂/FiO₂: Partial pressure of arterial oxygen/fraction of inspired oxygen. *comparison among the groups, Repeated Measures of ANOVA. There was no difference in PaO₂, PaCO₂, pH, SaO₂, [HCO₃⁻], base excess, lactate, and PaO₂/FiO₂ values among the groups (P = 0.299, 0.843, 0.632, 0.736, 0.685, 0.698, 0.027, and 0.268, respectively) (P < 0.016 indicates statistically significant difference among the groups). [†]within group comparisons, Repeated Measures of ANOVA. There is a significant difference in PaO₂, PaCO₂, pH, SaO₂, [HCO₃⁻], base excess and PaO₂/FiO₂ values within group analysis (P < 0.001 for all) (P < 0.05 indicates statistically significant difference within group comparisons).

Table 4. Lung Compliance, Airway Resistance, and Cumulative Morphine Consumption

	Group TIVA (n = 27)	Group D (n = 27)	Group S (n = 28)	P value
C (ml/cmH ₂ O)				0.176*
After intubation [†]	62.7 ± 19.4	66.7 ± 19.3	58.9 ± 15.8	
1st h of operation [†]	57.5 ± 13.4	62.2 ± 19.5	54.0 ± 14.0	
2nd h of operation [†]	53.8 ± 11.7	51.8 ± 11.0	47.4 ± 9.2	
P value [†]	< 0.001 [†]	< 0.001 [†]	< 0.001 [†]	
R _{aw} (cm/H ₂ O/L/s)				0.697*
After intubation [†]	9.8 ± 5.4	9.0 ± 1.7	9.4 ± 2.5	
1st h of operation [†]	9.5 ± 2.4	9.1 ± 2.0	9.9 ± 2.7	
2nd h of operation [†]	9.3 ± 2.2	8.9 ± 1.3	9.8 ± 1.6	
P value [†]	0.808	0.778	0.372	
Cumulative Morphine consumption ^{§,} (mg)	7.1 ± 1.1	6.7 ± 1.1	7.5 ± 1.0	0.030*

Values are presented as mean ± SD. C: Compliance, R_{aw}: Airway resistance. *: comparison among the groups, [†]: Repeated Measures of ANOVA, [§]: Kruskal Wallis, ^{||}: Bonferoni corrected Mann Whitney U. There is no difference in dynamic lung compliance, airway resistance and cumulative morphine consumption among the groups (P = 0.176, 0.679 and 0.030 respectively) (P < 0.016 indicates statistically significant difference among the groups). [†]: inter-group comparisons, repeated measures ANOVA. The dynamic lung compliance reduced at the 1st and 2nd h of operation than after intubation in all groups (P < 0.001 for all). No difference in airway resistance at any measurement intervals within the groups (P = 0.808 in the Group TIVA, P = 0.778 in the Group D, P = 0.372 in the Group S) (P < 0.05 indicates statistically significant difference within group comparisons).

perform a proper spirometry test at the postoperative 30 min, and one vomited following extubation, which led to hypoxemia. Two patients from Group S were excluded, since one was admitted to intensive care unit due to severe intraoperative bleeding and the other could not postoperatively perform a proper spirometry test (Fig. 1).

The study groups demonstrated significant similarity with respect to ASA physical status, gender, age, body weight, height, and body mass index. Duration of the surgery was longer in the Group D than Group S (P < 0.001, Table 1). The study groups were also similar with regard to their heart rate, blood pressures, and respiratory rates (P > 0.05).

We did not identify any statistically significant inter-group differences with respect to FEV₁ levels in each measurement

interval (P = 0.336). There was a statistically significant reduction in the FEV₁ levels at the postoperative 30 min timepoint compared to the baseline, following which its levels increased and was significantly higher at the postoperative 24 h than at the postoperative 30 min in all groups (P = 0.001, Table 2).

Although the baseline FVC values were lower in the Group S than Group TIVA and D, the difference was not statistically significant (P = 0.031, Table 2). The reduction in the postoperative 30 min FVC values demonstrated statistical significance than that at the baseline in all groups (P < 0.001 for the Group TIVA and Group D, 0.002 for the Group S). FVC values demonstrated a statistically significant increase at the postoperative 24 h than that at the postoperative 30 min in all groups (P < 0.001 for the Group TIVA and Group D, 0.002 for the Group S).

According to the data, we did not identify any particular inter-group differences with regard to the FEV₁/FVC, PaO₂, PaCO₂, pH, SaO₂, [HCO₃⁻], base excess, lactate, and PaO₂/FiO₂ (P = 0.104, 0.299, 0.843, 0.632, 0.736, 0.685, 0.698, 0.027, and 0.268, respectively) (Tables 2 and 3). We noted statistically significant differences in the outcomes of the intra-group analysis with regard to FEV₁/FVC values in Group S (P = 0.009) and PaO₂, PaCO₂, pH, SaO₂, [HCO₃⁻], base excess, and PaO₂/FiO₂ in all groups (P < 0.001 for all).

Dynamic lung compliance did not demonstrate any statistically significant difference at any measurement interval between the groups (P = 0.176). Reduction in the dynamic lung compliance levels was statistically significant at the 1st and 2nd hour of the operation than that after intubation in all groups (P < 0.001 for all, Table 4). Furthermore, there was no statistically significant difference with respect to airway resistance at any measurement intervals between (P = 0.679) and within the groups (P = 0.808 in the Group TIVA, P = 0.778 in the Group D, P = 0.372 in the Group S, Table 4).

Cumulative morphine consumption was not different among the groups (P = 0.030, Table 4). None of the patients needed additional analgesic at the postoperatively.

Discussion

Similar to the previous studies, we identified a reduction in FEV₁ and FVC values in the patients awakening from general anesthesia [13–15]. Considering the primary endpoint of the present study, we did not find any inter-group differences in FEV₁ values. Additionally, we also did not observe any differences in secondary endpoints (FVC, FEV₁/FVC, ABG analysis, compliance, and airway resistance) among the groups.

Inducing the general anesthesia reduced the functional residual capacity and increased airway resistance [16]. These changes may eventually lead to airway closure, atelectasis, or ventilation/perfusion mismatch [1]. Although preoxygenation with 100% O₂ helped to prevent hypoxemia in cases demonstrating difficulty in ventilation and intubation; additionally, rapid absorption of trapped gas within closed airways may lead to atelectasis [1,17]. Alternatively, World Health Organization and the United States Center of Disease Control have recently recommended the intraoperative and early post-operative use of 0.8 FiO₂ to prevent surgical site infections [18]. However, this recommendation triggered several discussions [19]. Despite these controversies pertaining to the prevention of absorption atelectasis, we administered 0.8 FiO₂ while inducing the anesthesia and 0.4 FiO₂ to maintain general anesthesia.

Tiefenthaler et al. [9] compared the effects of propofol and sevoflurane on postoperative lung function tests in patients undergoing lumbar disc surgery. Although they too did not identify

any differences with regard to the FEV₁ values, the reduction in the FVC values reported by them were significantly greater in the propofol group than the sevoflurane group. In their study, the patients were operated while being placed in a prone position, whereas in our study the patients were placed in a supine position. Different results may be attributed to the effect of surgical position on the ventilation/perfusion mismatch. Furthermore, they did not report any change in their primary endpoint (FEV₁) and only showed a difference in their secondary endpoint (FVC). This may be due to a type II bias. Additionally, Tiefenthaler et al. [9] have not disclosed their intraoperative ventilation strategy, which may have affected the study results. Kim et al. [11] compared the effects of propofol and desflurane on the postoperative respiratory functions of elderly patients undergoing knee surgery. Similar to our results, they did not observe any differences in the postoperative spirometry values. Ozdogan et al. [12] compared the effects of sevoflurane and desflurane on respiratory function in patients undergoing sleeve gastrectomy and did not report any significant differences. Although a study with animal subjects had reported that desflurane increased total lung resistance and decreased lung compliance, sevoflurane did not [20]. Similar to Ozdogan et al. [12], we did not detect any differences in postoperative respiratory function tests between sevoflurane and desflurane groups. Here, intraoperative dynamic lung compliance values were reduced in the 1st and 2nd hours instead of immediately after intubation. It is our understanding that it might have been better to titrate the positive airway pressure level according to lung compliance; however, we used 5 cmH₂O PEEP in all patients to avoid increased intracranial pressure. We did not observe any inter-group differences with regard to airway resistance. Although it has previously been demonstrated that sevoflurane caused airway relaxation while desflurane did not, we did not observe any significant differences between the groups [6–8]. Additionally, Zoremba et al. [10] compared the effects of propofol to desflurane on early postoperative lung functions in overweight patients and determined that propofol impaired early postoperative lung functions more than desflurane. Propofol decreased upper airway tone, unlike the volatile anesthetics [1], and this may be more significant in overweight patients [21].

In the present study, the PaO₂ levels were higher during the intraoperative period and after 30 min postoperatively than the baseline values and 24 h due to O₂ administration. The PaCO₂ levels were higher and pH values were lower after 30 min postoperative than the other sampling times due to early postoperative hypoventilation. Other statistically significant differences in terms of ABG analysis and PaO₂/FiO₂ were not clinically important.

In the present study, intraoperative analgesia was provided using remifentanyl and morphine administration. The nasal

phase of the endoscopic endonasal transsphenoidal pituitary surgery is extremely painful. Controlled hypotension was necessary during this phase of the surgery to prevent severe bleeding. Preoperative local anesthetic and adrenalin administration to the nasal mucosa of concha reduced the amount of bleeding, but systemic absorption of adrenalin facilitated the rise in blood pressure. We administered remifentanyl (max dose: 0.2 µg/kg/min) with morphine (max dose: 0.1 mg/kg) to provide analgesia during the nasal phase of the surgery. None of our patients needed additional postoperative analgesia. Cumulative morphine consumption was similar across the groups. Therefore, morphine induced hypoventilation could not impact our study results.

This study has a number of limitations, the study duration could have been longer and any surgical intervention at the nasal cavity may interfere with the spiograms. Additionally, we could not titrate the positive airway pressure level according to lung compliance. We used 5 cmH₂O PEEP in all patients to avoid raised intracranial pressure. Therefore, there was a remarkable reduction in the intraoperative dynamic compliance after intubation. It was our understanding that we could have better described the negative effects of anesthetic drugs on respiratory functions since intracranial surgery patients were already on a suboptimal ventilation with lower PEEP and without the utilization of recruitment maneuvers. Although the numerical values of the baseline FVC were lower in Group S than Groups TIVA and D, the difference was not statistically significant. Therefore, we believe that this may not have affected our study results.

In conclusion, we compared the effects of propofol, desflurane, and sevoflurane on respiratory functions in patients undergoing endoscopic endonasal transsphenoidal pituitary surgery during the postoperative 24 hours. We demonstrated that all three agents postoperatively reduced respiratory functions, without any significant differences among the drugs.

References

1. Hedenstierna G, Tokics L, Lundquist H, Andersson T, Strandberg A, Brismar B. Phrenic nerve stimulation during halothane anesthesia. Effects of atelectasis. *Anesthesiology* 1994; 80: 751-60.
2. Zhang XJ, Yu G, Wen XH, Lin ZC, Yang FQ, Zheng ZG, et al. Effect of propofol on twitch diaphragmatic pressure evoked by cervical magnetic stimulation in patients. *Br J Anaesth* 2009; 102: 61-4.
3. Domino KB, Borowec L, Alexander CM, Williams JJ, Chen L, Marshall C, et al. Influence of isoflurane on hypoxic pulmonary vasoconstriction in dogs. *Anesthesiology* 1986; 64: 423-9.
4. Loer SA, Scheeren TW, Tarnow J. Desflurane inhibits hypoxic pulmonary vasoconstriction in isolated rabbit lungs. *Anesthesiology* 1995; 83: 552-6.
5. Schwarzkopf K, Schreiber T, Preussler NP, Gaser E, Hüter L, Bauer R, et al. Lung perfusion, shunt fraction, and oxygenation during one-lung ventilation in pigs: the effects of desflurane, isoflurane, and propofol. *J Cardiothorac Vasc Anesth* 2003; 17: 73-5.
6. Prakash YS, Iyanoye A, Ay B, Sieck GC, Pabelick CM. Store-operated Ca²⁺ influx in airway smooth muscle: Interactions between volatile anesthetic and cyclic nucleotide effects. *Anesthesiology* 2006; 105: 976-83.
7. Goff MJ, Arain SR, Ficke DJ, Uhrich TD, Ebert TJ. Absence of bronchodilation during desflurane anesthesia: a comparison to sevoflurane

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Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

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- and thiopental. *Anesthesiology* 2000; 93: 404-8.
8. von Ungern-Sternberg BS, Saudan S, Petak F, Hantos Z, Habre W. Desflurane but not sevoflurane impairs airway and respiratory tissue mechanics in children with susceptible airways. *Anesthesiology* 2008; 108: 216-24.
 9. Tiefenthaler W, Pehboeck D, Hammerle E, Kavakebi P, Benzer A. Lung function after total intravenous anaesthesia or balanced anaesthesia with sevoflurane. *Br J Anaesth* 2011; 106: 272-6.
 10. Zoremba M, Dette F, Hunecke T, Eberhart L, Braunecker S, Wulf H. A comparison of desflurane versus propofol: the effects on early postoperative lung function in overweight patients. *Anesth Analg* 2011; 113: 63-9.
 11. Kim YS, Lim BG, Kim H, Kong MH, Lee IO. Effects of propofol or desflurane on post-operative spirometry in elderly after knee surgery: a double-blind randomised study. *Acta Anaesthesiol Scand* 2015; 59: 788-95.
 12. Ozdogan HK, Cetinkunar S, Karateke F, Cetinalp S, Celik M, Ozyazici S. The effects of sevoflurane and desflurane on the hemodynamics and respiratory functions in laparoscopic sleeve gastrectomy. *J Clin Anesth* 2016; 35: 441-5.
 13. Diamant ML, Palmer KN. Postoperative changes in gas tensions of arterial blood and in ventilatory function. *Lancet* 1966; 2: 180-2.
 14. Hedenstierna G, Löfström J. Effect of anaesthesia on respiratory function after major lower extremity surgery. A comparison between bupivacaine spinal analgesia with low-dose morphine and general anaesthesia. *Acta Anaesthesiol Scand* 1985; 29: 55-60.
 15. Sah HK, Akcil EF, Tunali Y, Vehid H, Dilmen OK. Efficacy of continuous positive airway pressure and incentive spirometry on respiratory functions during the postoperative period following supratentorial craniotomy: a prospective randomized controlled study. *J Clin Anesth* 2017; 42: 31-5.
 16. Hedenstierna G. Oxygen and anesthesia: what lung do we deliver to the post-operative ward? *Acta Anaesthesiol Scand* 2012; 56: 675-85.
 17. Edmark L, Kostova-Aherdan K, Enlund M, Hedenstierna G. Optimal oxygen concentration during induction of general anesthesia. *Anesthesiology* 2003; 98: 28-33.
 18. Allegranzi B, Zayed B, Bischoff P, Kubilay NZ, de Jonge S, de Vries F, et al. New WHO recommendations on intraoperative and postoperative measures for surgical site infection prevention: an evidence-based global perspective. *Lancet Infect Dis* 2016; 16: e288-303.
 19. Akca O, Ball L, Belda FJ, Biro P, Cortegiani A, Eden A, et al. WHO Needs High FiO₂? *Turk J Anaesthesiol Reanim* 2017; 45: 181-92.
 20. Satoh JI, Yamakage M, Kobayashi T, Tohse N, Watanabe H, Namiki A. Desflurane but not sevoflurane can increase lung resistance via tachykinin pathways. *Br J Anaesth* 2009; 102: 704-13.
 21. Eastwood PR, Platt PR, Shepherd K, Maddison K, Hillman DR. Collapsibility of the upper airway at different concentrations of propofol anesthesia. *Anesthesiology* 2005; 103: 470-7.